

Thermal Oxidative Stability of Various Vegetable Oils used for the Preparation of the Seasoned Laver *Pyropia* spp.

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Abstract

Seasoned laver *Pyropia* spp. is a traditional Korean seafood that has gained popularity worldwide because of its unique taste, texture, and health benefits. It is prepared by roasting a sheet of dried laver, to which vegetable oils have been applied, at an ultra-high temperature (UHT) of 300°C. Therefore, the oxidative stability of the oils is the most important factor in determining the shelf life of seasoned laver products. In this study, we investigated changes in the thermal oxidative stability of six major vegetable oils (sesame, perilla, sunflower, rice bran, canola, and olive) during the seasoned laver processing. The oxidation induction time of each oil from the seasoned laver products was decreased compared with the fresh oil. These results indicate that the UHT treatment (300°C, 10 s) induced thermal oxidation of the oils. Among the six seasoned laver oils, the induction times of olive (OL, 8.02 h) and sesame (SE, 5.31 h) oils were significantly higher than the other oils. The acid values (AVs) of OL and SE oils from the seasoned laver were 0.49 and 0.79, respectively. On the other hand, perilla oil had the overall worst thermal oxidative properties (induction time: 0.35 h, AV: 2.82). Our results provide useful information about seasoned laver products for researchers or manufactures.

Key words: *Pyropia* spp., Seasoned laver, Vegetable oil, Thermal oxidation, Oxidative stability

Introduction

Seasoned laver *Pyropia* spp., the Korean traditional seafood, is a unique seaweed product that is prepared by roasting a sheet of dried laver, to which vegetable oils have been applied, at an ultra-high temperature (UHT) of 300°C for about 10 s. Korean seasoned laver products have been exported to over 70 countries, posting a profit of USD 180 million in 2013 (Cho et al., 2009; Korea Custom Service, 2014), and were second place in the seafood category (Park et al., 2001; Cho et al., 2009). In recent years, this product has become more popular worldwide because of its special taste, compactness, texture, and health benefits.

Vegetable oils are the most important ingredient (30–40%, w/w) in the preparation of seasoned laver products (Lee, 1999; Jo et al., 1995). In commercial seasoned laver products, sesame and perilla oils are most frequently used. During the seasoned laver processing, UHT treatment at 300°C induces thermal oxidation of the vegetable oils (Jo et al., 1995). Owing to the high oil content and UHT treatment, the oxidative stability of the oil is the most important factor in determining the shelf life of seasoned laver products (Wang et al., 2010; Hasenhuettl and Wan, 1992).

The general shelf life of seasoned laver supplied in domes-

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tic grocery-retail is about 6–12 months (Lee, 1999; Cho et al., 2009). This short shelf life is becoming a major problem in establishing export of the seasoned laver. Therefore, studies on prolongation of the shelf life of seasoned laver based on inhibition of oil oxidation are a promising research field in the Korean seaweed industry. Although various vegetable oils, such as sesame and perilla oils, are used in the seasoned laver preparation, there are few studies on their thermal oxidation stability.

In the present study, we investigated changes in the thermal oxidative properties of six major vegetable oils (sesame, perilla, sunflower, rice bran, canola, and olive) during the seasoned laver processing. The oxidative stability of each oil was evaluated by determination of the oxidation induction time using a Rancimat instrument. Moreover, the acid value, peroxide value, *p*-anisidine value, total polar compounds, and antioxidant capacity of each oil were analyzed as thermal oxidative stability parameters.

Materials and Methods

Materials

Sunflower (SF), rice bran (RB), canola (CA), and olive (OL) oils were purchased from CJ Cheiljedang Co., Ltd. (Seoul, Korea). Sesame (SE) and perilla (PE) oils were purchased from Chamgoeul Co., Ltd. (Seoul, Korea). These oils were stored in a refrigerator until use in an experiment. Sheet-type dried laver (*Pyropia* spp.) was purchased from Korea Fishery Co., Ltd (Pyeongtaek, Korea). All reagents used were analytical grade and purchased from Sigma-Aldrich Co. (St. Louis, MO, USA).

Oil sample preparation

For preparation of the seasoned laver, a sheet of dried laver (2.5 g) was heated at 300°C for 10 s after applying oils (1.6 g). The seasoned laver oils were then isolated using a press machine.

Oxidative stability using a Rancimat method

The oxidative stability of the fresh and seasoned laver oil samples were evaluated by measuring the oxidation induction time using a Rancimat instrument (Metrohm CH series 743, MetrohmAG, Herisau, Switzerland). The oxidation process was monitored for a 3 g sample of oil using an air velocity of 20 L/h at 120°C. During the Rancimat test, volatile compounds, such as formic acid, were rapidly produced and carried by the steam of air to be collected in deionized water. The vapors were continually monitored by measuring the conductivity of the deionized water. The end point was determined in hours.

Measurement of acid and peroxide values

To evaluate the stability of oils against thermal oxidation, the acid values (AVs) of the oils were determined utilizing an AOCS official method (AOCS, 1990). In brief, 10 g of each oil was weighed into a 125 mL-flask and mixed with 100 mL of a diethyl ether:ethanol (1:1, v/v) solution with a couple of drops of 1% phenolphthalein in ethanol (v/v) for indicator. This solution was titrated with 0.1 N potassium hydroxide solution.

The peroxide values (POVs) were determined using an AOCS official method. In brief, 5 g of each sample was weighed into a 125 mL-flask and mixed with 35 mL of chloroform:acetic acid (2:3, v/v) and 1 mL of saturated potassium iodide solution and mixture was shaken vigorously for 2 min and stored under dark conditions for 10 min. Subsequently, 75 mL of distilled water was added to the mixture, which was then shaken again. For the titration, 1 mL of 5% starch solution as an indicator was added to the mixture, which was then titrated with 0.01 N sodium thiosulfate solution.

p-Anisidine value and total polar compounds

The *p*-anisidine value (*p*-AV) of each oil was measured according to the AOCS method Ti la-64. Approximately 100 mg of sample was mixed with 25 mL isooctane in a 125 mL flask and the absorbance of this mixture was measured at 350 nm using a UV-Vis spectrophotometer (Cary 50, Varian, Palo Alto, CA, USA). A 2.5 mL aliquot was mixed with 0.5 mL of 0.5% (w/v) *p*-anisidine in acetic acid and the absorbance was measured at 350 nm after 10 min. Each sample was assayed in triplicate. The *p*-AV was calculated as follows:

$$p\text{-AV} = 25 \times (1.2 \times A_2 - A_1)/W$$

where A1 is the absorbance at 350 nm before addition of *p*-anisidine, A2 is the absorbance at 350 nm after addition of *p*-anisidine, and W is the amount of sample (g).

The total polar compounds (TPCs) in each oil sample were determined by a Testo 270 cooking oil tester (Testo Ltd., Hampshire, UK) according to the manufacturer's operation guide. Each sample was maintained at 60°C for the measurement. The cooking oil tester provided the TPCs values by measuring changes in the dielectric constant of the oil samples, which is highly correlated with the amount of polar compounds in the sample. Each sample was assayed in triplicate.

Measurement of DPPH

The antioxidant capacity of each oil was determined by a method of Blois (1958) compared using a modified 2,2-diphenyl-1-picrylhydrazyl (DPPH) method. A portion of each oil (56 µL) was mixed with 5 mL of 0.10 mM DPPH in isooctane in a 50 mL tube and stored under dark conditions for 30

min. After standing, the absorbance of the each sample was measured at 517 nm using a UV-Vis spectrophotometer (Cary 50, Varian, Palo Alto, CA, USA). The free radical scavenging activity measured by the DPPH method was considered as the antioxidant capacity of each oil.

Measurement of color

The color levels of oils were determined using a colorimeter (CR-300, Minolta, Japan). Approximately 2 g of oil was transferred to a petri dish (diameter: 3.5 cm) and measured the Hunter color scale which L (lightness), a (redness), and b (yellowness). For the reference color white board were used and its color scales were L=93.79, a=-0.12, and b=-0.20, respectively.

Statistical analysis

The differences of experimental parameter in the samples were analyzed statistically using ANOVA and Tukey's multiple range test at $P < 0.05$ with the Statistical Analysis System software, Version 9.2 (SAS Institute, Cary, NC, USA).

Results and Discussion

Thermal oxidative stability

The thermal oxidative stabilities of six vegetable oils from seasoned laver products were evaluated using the Rancimat method. This is the most common test used to assess the oxidative stability of edible oils, which is expressed as the oxidation induction time (Farhoosh and Kafrani, 2011). The oxidation induction times of all the oils from the seasoned laver products were significantly decreased compared with the fresh oils (Fig. 1). These results mean that the UHT treatment used in the seasoned laver processing induces oxidation of the oils despite the short treatment time. Among the six seasoned laver oils, OL showed the highest thermal oxidative stability (induction time: 8.02 h). In previous studies (Chiou et al., 2009; Casal, 2010), OL is well known to have excellent oxidative stability. OL contains more monounsaturated (palmitic and oleic) acids than other vegetable oils and less polyunsaturated acids, such as linolenic acid, that is, OL is more stable than other vegetable oils and not easily broken down by thermal oxidation (Velasco and Dobarganes, 2002; Chiou et al., 2009). Moreover, there are a variety of polyphenols with antioxidant activity in OL that contribute to the oxidative stability (Pso-miadou and Tsimidou, 2002; Leonardis et al., 2013).

SE and PE oils are most frequently used for seasoned laver preparation owing to their excellent sensory properties. SE had the second most stable oxidation stability of the studied oils (oxidation induction time of 5.31 h) because SE contains natural antioxidants such as sesamin, sesamol, and sesamo-

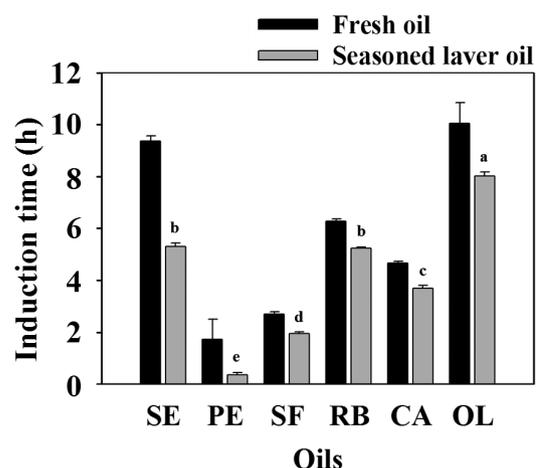


Fig. 1. Oxidative stabilities of fresh and seasoned laver *Pyropia* spp. oils. The oxidative stability is expressed as the oxidation induction time (h; mean \pm SD) determined by the Rancimat test. The superscripts (a–e) indicate significant differences ($P < 0.05$). SE, sesame oil; PE, perilla oil; SF, sunflower oil; RB, rice bran oil; CA, canola oil; OL, olive oil.

lin and is rich in tocopherols (Lee et al., 2008). In addition, SE has a high content of monounsaturated (oleic and linoleic) acids, which inhibit thermal oxidation (Bayder et al., 1999; Uzun et al., 2002). On the other hand, the induction time of PE (1.73 ± 0.77 h) showed the lowest oxidation stability among the tested oils, and the greatest decrease (79.7%) compared with the fresh oil. This result indicates that PE contains a high content (59%) of oxidation-susceptible linolenic acid (Wang, 2010; Zhao, 2012), which can induce rapid oxidation during UHT treatment.

AV and POV

AV and POV are important indexes of food quality for seasoned laver products. According to the Ministry of Food and Drug Safety (2011), the standard values of AV and POV in seasoned laver products are 4.0 mg KOH/g and 60.0 meq/kg, respectively. The AV indicates the level of free fatty acids in oils. Oils are oxidized by the action of oxygen or microorganisms during storage and the amount of free fatty acids are an indicator of the degree of oxidation (Kim et al., 2010). The AVs of the six vegetable oils are presented in Table 1. The AVs of OL, SE, RB, CA, SF, and PE oils from the seasoned laver products were 0.49, 0.79, 0.59, 0.11, 0.14, and 2.82 mg KOH/g, respectively. The AV of PE was significantly higher than those of the other oils, whereas CA had the lowest AV level ($P < 0.05$). This result is caused by the higher unsaturated fatty acid level in PE and is similar to the trend in induction times obtained using the Rancimat method.

The POV is determined using the peroxide content that is produced from oxidation of the oils (Kim et al., 2010). The

POVs of the six vegetables oils are shown in Table 1. The POVs of OL, SE, RB, CA, SF, and PE oils from the seasoned laver products were 11.26, 2.28, 5.42, 3.60, 5.61, and 4.78 meq/kg, respectively. The POV of OL was significantly higher than those of the other oils ($p < 0.05$), whereas SE had a significantly lower POV level. Even though OL has a lower polyunsaturated fatty acid content than the other oils, the POV of OL was higher compared to those of the other oils owing to its lower content of antioxidant components.

p-AV and TPC

The p-AV indicates the amount of aldehyde and ketone compounds produced during the thermal oxidation of oils (Min, 1984). The p-AV and TPCs of the vegetable oils are presented in Table 2. The p-AVs of OL, SE, RB, CA, SF, and PE oils from the seasoned laver products were 10.31, 7.96, 20.64, 6.59, 8.92, and 14.15, respectively. The p-AV of RB was significantly higher than those of the other oils, whereas CA had a significantly lower p-AV level ($P < 0.05$).

The TPC indicates the level of oxidative breakdown products present in an oil. The TPC includes components, such as free fatty acids, peroxides, aldehydes, acids, alcohols, and ketones, obtained from degradation of the oils. Higher TPC levels indicate poorer quality oils, and the literature notes that TPC levels greater than 25% mean that the oil should be discarded (Sebastian et al., 2014). The TPCs of OL, SE, RB, CA, SF, and PE oils from the seasoned laver products were 4.5, 9.5, 10.5, 6.8, 8.0, and 19.3%, respectively (Table 2). The TPC of PE was significantly higher than those of the other oils, whereas OL had a significantly lower TPC level ($P < 0.05$).

Table 1. Acid value (AV) and peroxide value (POV) of fresh and seasoned laver *Pyropia* spp. oils

Oils	AV (mg KOH/g)	POV (meq/kg)
SE Fresh oil	0.78 ± 0.03	0.10 ± 0.02
SE Seasoned laver oil	0.79 ± 0.01 ^b	2.28 ± 0.09 ^e
PE Fresh oil	3.15 ± 0.11	1.20 ± 0.12
PE Seasoned laver oil	2.82 ± 0.43 ^a	4.78 ± 0.23 ^c
SF Fresh oil	0.04 ± 0.02	3.68 ± 0.60
SF Seasoned laver oil	0.14 ± 0.01 ^{cd}	5.61 ± 0.09 ^b
RB Fresh oil	0.53 ± 0.07	1.93 ± 0.48
RB Seasoned laver oil	0.59 ± 0.04 ^{bc}	5.42 ± 0.07 ^{bc}
CA Fresh oil	0.07 ± 0.02	1.87 ± 0.41
CA Seasoned laver oil	0.11 ± 0.01 ^d	3.60 ± 0.47 ^d
OL Fresh oil	0.43 ± 0.01	9.17 ± 0.40
OL Seasoned laver oil	0.49 ± 0.02 ^{bcd}	11.26 ± 0.41 ^a

The results are expressed as mean ± SD (n = 3). The superscript letters (a-e) indicate significant differences by Tukey's multiple range test ($P < 0.05$). SE, sesame oil; PE, perilla oil; SF, sunflower oil; RB, rice bran oil; CA, canola oil; OL, olive oil.

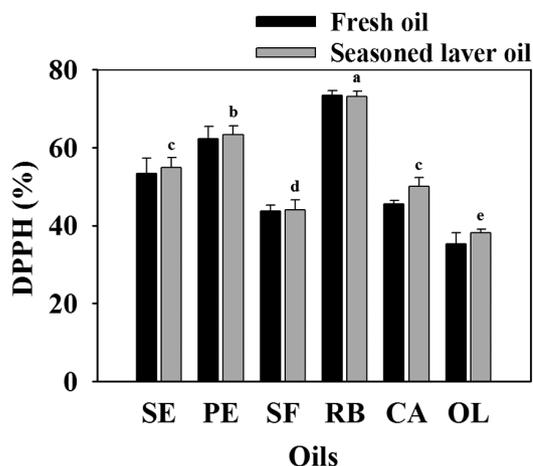


Fig. 2. Antioxidant capacities (DPPH) of fresh and seasoned laver *Pyropia* spp. oils. The results are expressed as mean ± SD (n = 3). The superscripts (a-e) indicate significant differences ($P < 0.05$). SE, sesame oil; PE, perilla oil; SF, sunflower oil; RB, rice bran oil; CA, canola oil; OL, olive oil.

Antioxidant capacity

The DPPH assay determines the free radical scavenging capacity as a measure of the antioxidant capacity in samples. DPPH has a stable radical with a maximum absorbance at 517 nm, and the absorbance is decreased by the addition of an electron or hydrogen radical. The antioxidant capacities of the vegetable oils are shown in Fig 2. The free radical scavenging capacities of the seasoned SE, PE, SF, BR, CA, and OL were 54.88 ± 2.46, 63.26 ± 2.23, 43.96 ± 2.59, 73.08 ± 1.35,

Table 2. p-Anisidine value (p-AV) and total polar compounds (TPC) of fresh and seasoned laver *Pyropia* spp. oils

Oils	p-AV	TPC (%)
SE Fresh oil	6.42 ± 1.76	8.5 ± 0.1
SE Seasoned laver oil	7.96 ± 1.02 ^{cd}	9.5 ± 0.1 ^c
PE Fresh oil	12.59 ± 2.09	18.3 ± 0.3
PE Seasoned laver oil	14.15 ± 1.12 ^b	19.3 ± 0.1 ^a
SF Fresh oil	6.73 ± 1.85	7.3 ± 0.3
SF Seasoned laver oil	8.92 ± 0.10 ^{cd}	8.0 ± 0.1 ^d
RB Fresh oil	20.20 ± 2.25	9.8 ± 0.3
RB Seasoned laver oil	20.64 ± 1.20 ^a	10.5 ± 0.1 ^b
CA Fresh oil	5.95 ± 1.50	6.5 ± 0.1
CA Seasoned laver oil	6.59 ± 1.13 ^d	6.8 ± 0.3 ^e
OL Fresh oil	8.80 ± 0.89	2.8 ± 0.3
OL Seasoned laver oil	10.31 ± 1.15 ^c	4.5 ± 0.1 ^f

The results are expressed as mean ± SD (n = 3). The superscript letters (a-d) indicate significant differences by Tukey's multiple range test ($P < 0.05$). SE, sesame oil; PE, perilla oil; SF, sunflower oil; RB, rice bran oil; CA, canola oil; OL, olive oil.

Table 3. Colorimetry of fresh and seasoned laver *Pyropia* spp. oils

	Oils	Colorimetry		
		L	a	b
SE	Fresh oil	31.25 ± 0.01	-0.20 ± 0.01	+3.27 ± 0.01
	Seasoned laver oil	30.58 ± 0.01 ^c	-0.91 ± 0.01 ^b	+4.61 ± 0.02 ^e
PE	Fresh oil	31.32 ± 0.01	-0.33 ± 0.03	+3.98 ± 0.01
	Seasoned laver oil	27.80 ± 0.12 ^e	-0.85 ± 0.01 ^a	+5.05 ± 0.01 ^c
SF	Fresh oil	32.77 ± 0.03	+0.33 ± 0.02	+0.13 ± 0.01
	Seasoned laver oil	29.97 ± 0.01 ^d	-1.60 ± 0.03 ^d	+5.00 ± 0.01 ^d
RB	Fresh oil	32.66 ± 0.05	+0.04 ± 0.03	+0.86 ± 0.01
	Seasoned laver oil	31.44 ± 0.01 ^a	-1.77 ± 0.01 ^e	+5.27 ± 0.02 ^b
CA	Fresh oil	32.61 ± 0.02	+0.30 ± 0.04	+0.29 ± 0.02
	Seasoned laver oil	31.56 ± 0.01 ^a	-1.34 ± 0.02 ^e	+4.25 ± 0.01 ^f
OL	Fresh oil	32.41 ± 0.34	-0.90 ± 0.02	+3.74 ± 0.01
	Seasoned laver oil	30.87 ± 0.01 ^b	-1.75 ± 0.02 ^e	+5.87 ± 0.01 ^a

The results are expressed as mean ± SD (n = 3). The superscript letters (a-e) indicate significant differences by Tukey's multiple range test ($P < 0.05$). SE, sesame oil; PE, perilla oil; SF, sunflower oil; RB, rice bran oil; CA, canola oil; OL, olive oil; L, lightness; a, redness; b, yellowness.

50.01 ± 2.31, and 38.17 ± 0.81%, respectively. BR showed the highest antioxidant capacity, whereas OL had the lowest ($P < 0.05$). Similar to the POV results, OL showed a lower antioxidant capacity compared with the other oils. Silva and others (2010) noted that the radical-scavenging activity measured by the DPPH assay is correlated with tocopherol content in edible oils. Tocopherol has four isomers: α -, β -, γ -, and δ -tocopherols, which have the following order of antioxidant capacity: δ - > γ - > β - > α -tocopherol. BR, PE, and SE are rich in δ - and γ -tocopherols, and BR especially is an excellent source of tocotrienols (Eitenmiller and Lee 2004).

Color

The color of an oil is one of the most important physical indicators for the determination of oil quality. The color scales of the vegetable oils were measured by a colorimeter and are shown in Table 3. The L- (lightness) values of the oils ranged from 27.80 ± 0.12 in PE to 31.56 ± 0.01 in CA; RB and CA showed significantly higher L-values ($P < 0.05$). PE had a significantly higher a- (redness) value ($P < 0.05$). Moreover, OL had a significantly lower a-value and higher b- (yellowness) value (5.87 ± 0.01). CA showed the highest L-value (31.56 ± 0.01) and lowest b-value (4.25 ± 0.01), and RB showed the high b-value (5.27 ± 0.02) but lowest a-value (-1.77 ± 0.01) ($P < 0.05$).

In summary, OL and SE oils showed excellent oxidative stability under UHT conditions in terms of oxidation induction time, and their AV and POV were lower than the standard values of the Ministry of Food and Drug Safety. The oil used in the production of commercial seasoned laver products is a mixture of two or three vegetable oils. Therefore, studies on

the influence of the formulation ratio of the oil mixture are needed in the future. Our other study (Cho et al., 2015), which deal the effects of the blending ratio of corn, sesame, and perilla oils on the oxidation and sensory quality of seasoned laver, may be good information.

Acknowledgments

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References

- AOCS. AOCS official and tentative methods of the American oil chemists' society (4th ed). Champaign, IL: AOCS Press (1990).
- Baydar H, Marquard R and Turgut I. 1999. Pure line selection for improved yield, oil content and different fatty acid composition of sesame, *Sesamum indicum*. Plant Breeding 118, 462-464.
- Blois MS. 1958. Antioxidant determinations by the use of a stable free radical. Nature 181, 1199-1200.
- Casal S, Malheiro R, Sendas A, Oliveira BPP and Pereira JA. 2010. Olive oil stability under deep-frying conditions. Food Chem Toxicol 48, 2972-2979.
- Chiou A, Kalogeropoulos N, Salta FN, Efstathiou P and Andrikopoulos NK. 2009. Pan-frying of French fries in three different edible oils enriched with olive leaf extract: Oxidative stability and fate of microconstituents. LWT – Food Sci Technol 42, 1090-1097.
- Cho S, Kim BM, Han KJ, Seo HY, Han YN, Yang EH and Kim DS. 2009. Current status of the domestic processed laver market and

- manufacturers. *Food Sci Indus* 42, 57-70.
- Cho S, Kim J, Yoon M, Yang H, Um MY, Park J, Park EJ, Yoo H, Baek J, Jo J. 2015. Monitoring and optimization of the effects of the blending ratio of corn, sesame, and perilla oils on the oxidation and sensory quality of seasoned laver *Pyropia* spp. *Fish Aquat Sci* xx, xx-xx.
- Eitenmiller RR and Lee J. 2004. Vitamin E: Food Chemistry, Composition, and Analysis. New York, NY: Marcel Dekker Inc.
- Farhoosh R and Kafrani MHT. 2011. Simultaneous monitoring of the conventional qualitative indicator during frying of sunflower oil. *Food Chem* 125, 209-213.
- Hasenhuettle GL and Wan PJ. 1992. Temperature effects on the determination of oxidative stability with the metrohm rancimat. *JAOCS* 69, 525-527.
- Jo KS, Kim JH and Shin HS. 1995. Effect of storage condition on the oxidative stability of lipid in roasted and roasted-seasoned laver (*Porphyra tenera*). *Kor J Food Sci Technol* 27, 902-908.
- Kim YS, Kim R, Na MS and Choi DB. 2010. Effect of extraction process on the physicochemical characteristics of seed oil *Camellia sinensis*. *Appl Chemr Engin* 21, 148-153.
- Korea Custom Service. 2014. Statistical Database for Seasoned laver Export. Retrieved from <http://www.customs.go.kr/> on 2014.
- Lee J, Lee Y and Choe E. 2008. Effect of sesamol, sesamin, and sesamol extracted from roasted sesame oil on the thermal oxidation of methyl linolate. *LWT – Food Sci Technol* 41, 1871-1875.
- Lee SK. 1999. Effect of packing on storage stability and chlorophyll contents of dried, roasted and roasted-seasoned laver during storage. *J Fd Hyg Safety* 14, 164-139.
- Leonardis AD, Angelico R, Macciola V and Ceglie A. 2013. Effects of polyphenol enzymatic-oxidation on the oxidative stability of virgin olive oil. *Food Res Int* 54, 2001-2007.
- Min DB. 1984. Lipid oxidation of edible oil. In: *Food Lipids*. p 283-296. New York, NY, USA.
- Park BH, Choi HK and Cho HS. 2001. A study on the oxidative stability and quality characteristics of *Kimbugak* made of aqueous green tea. *J Kor Soc Food Sci Nutr* 30, 567-564.
- Psomiadou E and Tsimidou M. 2002. Stability of Virgin Olive Oil. 1. Autoxidation Studies. *J Agric Food Chem* 50, 716-721.
- Sebastian A, Ghazani SM and Marangoni AG. 2014. Quality and safety of frying oils used in restaurants. *Food Res Int* 64, 420-423.
- Silva L, Pinto J, Carrola J and Paiva-Martins F. 2010. Oxidative stability of olive oil after food processing and comparison with other vegetable oils. *Food Chem* 121, 1177-1187.
- Uzun B, Ülger S and Çağirgan Mİ. 2002. Comparison of determination and Indeterminate types of sesame for oil content and fatty acid composition. *Turk J Agric For* 26, 269-274.
- Velasco J and Dobarganes C. 2002. Oxidation stability of virgin olive oil. *Eur J Lipid Sci Technol* 104, 661-676.
- Wang Y, Zhao M, Tang S, Song K, Han X and Ou S. 2010. Evaluation of the oxidative stability of diacylglycerol-enriched soybean oil and palm olein under rancimat-accelerated oxidation conditions. *J Am Oil Chem* 87, 483-491.
- Zhao TT, Hong SI, Lee J, Lee JS, and Kim IH. 2012. Impact of roasting on the chemical composition and oxidative stability of perilla oil. *J Food Sci* 77, C1273-C1278.